



ichthyologica, the aquarium journal

April-June 1967

Volume XXXIX

Number 2

One Dollar

CONTENTS

- EDITORIAL — OPPORTUNITIES FOR ICHTHYOLOGICAL MUTUALISM
Martin R. Brittan 55
- THE FUNCTIONAL SIGNIFICANCE OF THE SPLIT-HEAD COLOR
PATTERN AS EXEMPLIFIED IN A LEAF FISH, *POLYCENTRUS*
SCHOMBURGKII
George W. Barlow 57
- A RARE HAWKFISH *OXYCIRRHITES TYPUS* BLEEKER FOUND
IN HAWAII
Robert A. Morris and Donald E. Morris 71
- SULFAMERAZINE RESIDUES IN TROUT TISSUES
Roger Lee Herman and Paul E. Degurse 73
- THE DEATH VALLEY FISHES — AN ENDANGERED FAUNA
Martin R. Brittan 81

EDITORIAL

OPPORTUNITIES FOR
ICHTHYOLOGICAL MUTUALISM

MARTIN R. BRITTON

Over recent years I have received and read many bulletins put out by local aquarium societies. Most were dittoed or mimeographed and devoted to society news and notices, but many contained articles on the keeping and breeding of aquarium fishes. Some of this material was worthy of further attention by "professionals". Many of the contributors, almost all amateur aquarists, write poorly, but they are often capable people with intimate knowledge of certain fishes not possessed by ichthyologists. Much of the work goes to waste because the writers lack the training and acumen to fully appreciate and exploit their opportunities and because their hobbyist readers lack the same characteristics. As much goes unnoticed because far too many ichthyologists are too busy or too snobbish to be bothered with amateurs. Once in awhile a young aquarist develops into an ichthyologist, or a young ichthyologist begins to keep live fishes in connection with some scientific problem, and both fields consequently benefit enormously. But, in totality, this is, unfortunately, not anywhere near common enough. One group is not much involved in the problems of the other, and through lack of communication much is lost.

Many an ichthyologist looks down his nose at "naive and undisciplined amateurs", and many an aquarist looks up in intimidated awe or impatient boredom at the "impractical" expert.

What all this is leading up to is a plea: Ichthyologists should not overlook the books, clubs, and genuine information which may be supplied by hobbyists or dealers who keep and breed fishes, especially those fishes that are rare, unusual, or thought to be "difficult". Botanists have been willing to learn from, and quick to give credit to, the plant breeder, agriculturalist, and even the green-thumb garden-

EDITOR—Dr. Martin R. Britton, Sacramento State College, Sacramento 18, Calif.
EDITORIAL BOARD—Dr. Jacques Géry, Station Biologique, Les Eyzies, Dordogne, France; Dr. Leonard P. Schultz, Smithsonian Institution, Washington, D.C. ASSOCIATE EDITOR—Michael Reed, T.F.H. Publications, Inc. PRESIDENT—Dr. Herbert R. Axelrod, T.F.H. Publications, Inc., 245 Connelison Ave., Jersey City, N.J.

RATES: \$1.00 per copy in the U.S., \$1.50 per copy in Canada or foreign, \$10.00 for 12 issue subscription but special student rate of \$5.00 is also available. Foreign subscriptions are \$20 per year additional.

Second Class Postage Paid at Jersey City, New Jersey. Published quarterly by T.F.H. Publications, Inc. at 245 Connelison Avenue, Jersey City, N. J. 07310. Printed in U.S.A.

er (and a lot of aquarists have the watery equivalent of a green thumb). Aquarists, in turn, should learn to write down or tape-record their observations, and, more than that, do it systematically and accurately. If they do not, facts worthy of being recorded and passed on are simply forgotten and lost either to the hobby or science. In publishing their results, even in mimeographed publications, truly pertinent information should be given. An amateur can write interestingly without being cute or coy, distractingly breezy or chatty, long-winded or irrelevant. Information can be put in a form (with the help of a better writer, if necessary) such that a serious scientist, not to mention a scientist, will not gag on it!

There can be many dividends in increased communication and cooperation. For example, a key group in the work of fish behaviorists is, and has been for a long time, the family Cichlidae. Almost all species in this group were imported for and are still supplied by the aquarium trade. The earliest information about the group was gleaned not by ethologists but by aquarists and professional breeders. Behaviorists please note — there are several dozen other families commonly kept by aquarists.

On the other side of the coin, it should be pointed out that most of the really technical problems were, or ultimately will be, solved by experts, beginning with the fact that without the ichthyologist the aquarist would not even know what species of fishes he had in his tanks. It is, however, the aquarist who is most likely to appreciate his debt and his opportunity to learn. The ichthyologist surely has opportunities potentially as great.

SMITHSONIAN INSTITUTION
Washington, D. C. 20560

The Smithsonian Institution and T.F.H. Publications, Inc., are pleased to announce the publication of a reprint, including the color plates, of the Philippine Bureau of Science three Monographs on Philippine Fishes: No. 1, Jordan and Richardson's Checklist, 1909; No. 23, A. W. Herre's Gobies, 1927; and No. 24, Mammillari's Poeciliidae, 1927. These rare historical works are available in a clothbound volume for \$5.50.

Two earlier numbers in this reprint series are: Jordan and Evermann's "The Fishes of North and Middle America," U. S. Nat. Mus. Bull. 47, Vols. 1-4, 1900-1903, \$23.00; and Smith's "The Freshwater Fishes of Siam or Thailand," U. S. Nat. Mus. Bull. 188, 1945, \$2.00.

Orders for these books, accompanied by remittance (prepaid) should be addressed to:

Publications Distribution Section
Editorial and Publications Division
Smithsonian Institution
Washington, D. C. 20560

THE FUNCTIONAL SIGNIFICANCE OF THE SPLIT-HEAD
COLOR PATTERN AS EXEMPLIFIED IN A LEAF FISH,
POLYCENTRUS SCHOMBURGKII

GEORGE W. BARLOW

Department of Zoology
and
Museum of Vertebrate Zoology
University of California
Berkeley, California

It is the habit of illustrators of fishes to present their subjects as seen from the side. This is reasonable because fishes, being typically compressed, are most distinctive viewed in that way. Yet many fishes have markings on their heads that can only be appreciated in an adaptive context when seen from the front. Baerends and Baerends-van Roon (1950) created some awareness of this when they studied the hostile displays of cichlid fishes. They illustrated (p. 43, fig. 14) the frontal displays of certain species as they would be seen by their opponents, facing with the opercula extended; then the significance of the opercular flaps as large pseudo-eyes was fully appreciated. In a similar fashion, Abel (1964) figured the masks on the fronts of the faces of various male *Moenia*.

Many other special signals would be discovered if more fishes were viewed from the front. As an example in passing, certain surf perches (*Limniscidae*) have white lower lips. When seen from the side this feature is hardly noticed. But when a school turns and faces a diver the effect is dramatic, his eye being deflected to the many white spots in the water.

The focus of this article, however, is on the striking interorbital stripe as exemplified by a South American leaf fish, *Polycentrus schomburgkii* Müller and Troschel. An argument will be advanced for its function. This argument will be extended to other fishes evincing similar color patterns. Finally, some exceptions will be described.

Acknowledgments: I would like to take this opportunity to thank the people at the meeting of the American Society of Ichthyologists and Herpetologists who responded to enthusiastically (see below). For a stimulating discussion of the paper presented there I am particularly grateful to John Magnuson and Henry Föllmer. Edmund S. Hobson generously made available his knowledge and photographs garnered from underwater observations. Karl Linn offered supplemental observations on the feeding behavior of another leaf fish; Thomas Frazetta, John Mertz, and Peter Staller read the manuscript.

The basic observations were made under the auspices of a post-doctoral fellowship (MF 8244) from the National Institute of Mental Health. This was undertaken at the Max-Planck-Institut für Verhaltensphysiologie; I am eternally indebted to Konrad Z. Lorenz who made this possible, and who inspired me to learn about the behavior of animals. The writing itself was done under the support of a grant (GB 2210) from the National Science Foundation.

MATERIALS AND METHODS: The conditions under which the *Polycentrus* were kept are of little consequence to the behavior to be described here. Suffice it to say that the fish were kept in well-planted aquaria with conditions of water, temperature, and light appropriate to their natural environment. Details can be found in another article, one dealing with the social behavior of *Polycentrus* (Barlow 1967).

This paper was presented at the annual national meeting of the American Society of Ichthyologists and Herpetologists, during June, 1965, in Lawrence, Kansas. Taking advantage of the assembled body of knowledge present in the audience, I invited further examples of the kind of color pattern to be discussed in what follows (designated *A.S.I.H.*). For some time thereafter I continued to receive comments through the mail from ichthyologists who had heard about the paper; these will be marked *pers. comm.* Recently I observed several examples at the Steinhart Aquarium in San Francisco.

DIET: For approximately three months the diet of the *Polycentrus* had consisted largely of small amphipods, tubificid worms, and insect larvae. There was nothing remarkable about the feeding behavior during that time. The fish simply swam to the object to be eaten, paused, fixed on it, and engulfed it.

Then the fish were fed a school of young cichlids, *Europletis maculata* (Bloch). These small cichlids were difficult for the *Polycentrus* to catch by comparison with their previous food items. The cichlids hid among the plants, and fled and dodged artfully when approached by the *Polycentrus*.

For the first time I observed what is evidently the full feeding response of *Polycentrus*. An idealized account follows, but it is one based on repeated observations of feeding behavior.

When hunting, *Polycentrus* drifts slowly toward its intended prey. The only moving parts of its body are the lobes of the dorsal and anal fins and the pectoral fins; occasionally the caudal fin may also contribute to the locomotion. All of these structures are transparent. As the *Polycentrus* glides toward the prey the median fins and the paired pelvic fins become fully extended. A slight partial flexing and extending of the spines of the dorsal fin can sometimes be seen. When about one to two body lengths away from its prey, the fish pauses. A slight lateral curvature forms in the region of the caudal peduncle, imparting what I have termed a sickle-shape

in another article (Barlow 1967). The *Polycentrus* then darts forward, engulfing the prey if successful. The lunge finishes in a sharp turn to one side, starting at about the point where the prey is grasped. Details of the strike cannot be observed because of its extreme speed. But the mouth is apparently extended forward while the buccal cavity is expanded. In effect, the fish projects a tube toward the prey and sucks it in.

A remarkable aspect of the behavior of *Polycentrus* while hunting small fishes is the distinctive color pattern that appears. Both sexes show the color change, although the general color of the female does not become as dark as does that of the male; he often becomes nearly black while she becomes dark brown. Both sexes may show various white and black spots on this dark background (Fig. 1). The head in particular becomes nearly



Fig. 1.—Side view of a feeding *Polycentrus schomburgkii* showing the spotted pattern, the darker top of the head, and the inconspicuousness of the interorbital stripe seen from this angle. Photo by the author.

black, although the heavy black markings that run through it are still apparent. In juveniles the body may remain pale while the head darkens.

The singular feature of the color change is that which takes place on the top of the head. A cream-colored stripe develops, running from the junction of the premaxillaries, back between the eyes, and up the nape, ending at the insertion of the dorsal fin. It is narrowest, less than one eye diameter, on the snout, widening to a little more than an eye diameter on the nape. In frontal view (Fig. 2) the white stripe is continued onto the tips of the dorsal spines. The leading edges of the pelvic fins are also creamy white.

Thus seen from in front, from the prey's point of view (Fig. 3), *Polycoerus* gives the appearance of a vertical black ellipse, split by a white stripe running from its middle up to its top, and having two thin white stripes leading the pelvic fins.

Discussion: More than any other single effort, Cott's book (1940) on adaptive coloration has persuaded biologists to accept the principle of disruptive coloration in animal camouflage. His arguments of interest to us on this point can be summarized in a few lines: "It is this continuity of surface, bounded by a specific contour or outline, which enables us to recognize any object with whose shape we are familiar." (p. 48). So if *Polycoerus* is to minimize detection by its prey, it must break up its outline in some way.



Fig. 2.—Front view of a lurking *Polycoerus ichangensis* showing the extension of the white stripe onto the edge of the dorsal fin. Photo by the author.



Fig. 3.—Front view of a lurking *Polycoerus ichangensis* moments before engulfing a young *Daphnia mendotae* (slightly out of focus, directly before the mouth of the *Polycoerus*). Photo by the author.

Again, as Cott put it on page 52, "Broadly speaking . . . we may say that white marks on dark animals living in dark surroundings (such as forests) . . . will be the most effective in breaking up the continuity of their surface, and in making by contrast tall-tale half-tones of surface structure and modelling." On pages 46 to 49 Cott discussed a frog (Plate 9) that has a prominent mid-dorsal stripe, in many respects the functional counterpart of the interorbital stripe in *Polycoerus*. The emphatic stripe was said to achieve its effect in three ways: (1) it stands out, detaching the eye from the profile of the frog; (2) this strong incidence of color tends to flatten the half-tones by which the frog is recognized; and (3) the stripe bisects the frog—two half frogs do not resemble a frog.

In Cott's logic, the interorbital stripe of *Polycoerus* serves to break up its characteristic frontal profile. Evidently it is an adaptation to reduce detection while slowly approaching its prey.

In addition to the frog, Cott also discussed some insects (p. 79) that employ median stripes for disruptive effects. (His comments here about coincident disruptive patterns may also be extended to *Polycoerus* but need not concern us.) In these instances there is no suggestion of camouflage for predation, but just for protection.

For the sake of economy the discussion should be confined to fishes, yet one instance among mammals is especially worth noting. Charles Long drew my attention to the case of the badger. It is a predaceous burrower (Richardson 1929) with a split-head color pattern, and one that may lie in ambush for ground squirrels (Balph 1961).

Illustrations of fishes with disruptive stripes running up the interorbital space are difficult to find, but are not rare. Not only are fishes conventionally shown in side view, but they are usually drawn from dead specimens. If the pattern is seen only in the live fish, as in *Polycentrus*, even a figure depicting the frontal aspect will be of no avail. The grass pickerel (*Esox americanus vermiculatus* LeSueur) bears a permanent split-head pattern (Crossman 1966; pers. observ.), as must fishes that retain the interorbital stripe as museum material, e.g., *Acanthoplesiops indicus* (Day) (Smith 1950, fig. 409).

Longley (in Longley and Hildebrand 1941) photographed in nature two species with disruptive interorbital stripes, a trumpet fish *Aulostomus maculatus* Valenciennes, and a Nassau grouper *Epinephelus striatus* (Poey). An unidentified marine cel also has such an interorbital stripe in life (Roughly 1951, pl. 40) as does the cleaner wrasse *Labroides dimidiatus* (Clupaty 1963, fig. 3).

The presentation of this paper at the meeting of the American Society of Ichthyologists and Herpetologists stimulated the following further examples: The split-head pattern is found in the pomacentrid fish *Amphiprion akallopisus* Bleeker, and in some long-snouted chaetodontid fishes (L. P. Woods). It has also been seen (D. P. DeSilva) in the tripletail, *Lobotes surinamensis* (Bloch). Further examples are found in celpouts of the genus *Lycodes* occurring in the Bering Sea (N. Wilomovsky), in at least three gobioid fishes (J. C. Briggs), and in some gobies from the Philippine Islands (G. S. Myers). Preserved specimens of a sunfish, *Pomoxis nigromaculatus* (LeSueur), have been found with a ". . . distinct brown line extending from the gular region up across the snout onto the dorsal spines. This bisecting line is not characteristic of most of the members of this species . . ." (W. H. Walker, Jr., A.S.I.H. and pers. comm.). Finally, isospondylous fishes of the genus *Dixonina* lack the split-head color pattern but do have a conspicuous pale spot on the tip of the snout which may serve a similar end (F. H. Berry).

A visit to the Steinhart Aquarium in San Francisco resulted in several variations on the same theme. In the large squirrel fish *Holocentrus spinifer* (Forskål), the forehead is lighter red than the rest of the fish and is split by a brown-red line running from the mouth to just behind the interorbital space. The split-head effect is extremely pronounced in the lobotid fish *Datnioides microlepis* Bleeker; the leading edges of the pelvic fins are white, the posterior portions black. Adults of the serranid fish *Grammistes sex-*

April-June 1967

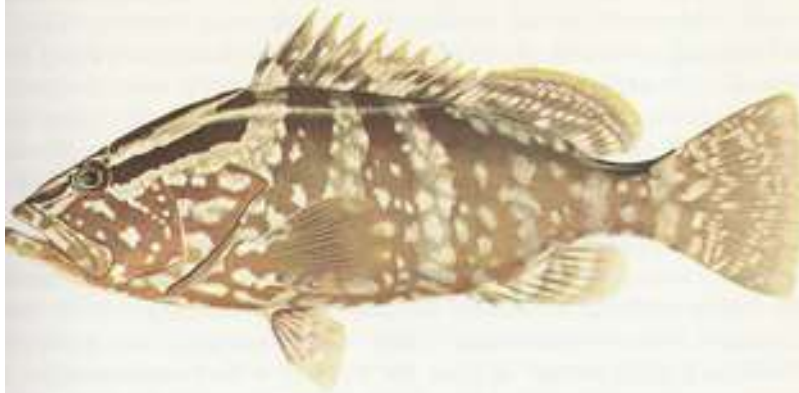


Fig. 4—A Nassau grouper *Epinephelus striatus*. Photo courtesy T.F.H. Pub.



Fig. 5—*Datniodes microlepis*. Photo by Peter Tsong.

linatus (Thunberg) are black with many white stripes and lack an interorbital stripe; it is present in the younger specimens, however. A reef-dwelling jack, *Gomus* sp., has a thin but distinct median stripe starting just above the eyes and running to the top of the head; I tried to view this species head on, but the fish always dropped down and looked slightly up at me, which placed the stripe in the middle of the obliquely-presented frontal profile. Two striking examples were seen among the anemone fishes of the genus *Amphiprion*: In both species a white middorsal stripe runs the length of the orange fish; in *A. akallopisos* Bleeker the stripe starts at the mouth, but in *A. posidonion* Bleeker it starts further back at the interorbital space. The stripe bisecting the face of the long-nosed butterfly fish *Chaetodon rostratus* (Linnaeus) is noteworthy for its border; the face is clear white and the stripe is gold, but set off from the white by a dark brown margin. A short but distinct split-head stripe was also observed in a pipefish *Syngnathus griseobasus* (Ayres), a blipfish *Gibbomia elegans* (Cooper), a file fish *Monacanthus* sp., and in an agonid fish. Among several eel-benties only one species, the pholid *Apoecilichthys flaccida* Girard, had a split-head pattern, a pronounced thin white line running middorsally from the snout to the tail; the same pattern, however, was noted in another pholid, *Xoropus fauorum* (Jordan & Gilbert), seen at the Bodega Marine Laboratory of the University of California.

In most of the cases just cited the fishes are thought to be predaceous, but useful observations are not readily available. In the following some examples are given where the split-head pattern has been seen, together with observations on the feeding habits of the animals.

The feeding behavior and color patterns of another leaf fish, and a closely related one, *Monoacanthus polyacanthus* Heckel, has been described by Cott (1940: 311-313, fig. 63). Cott dwelled on the appearance of the side of this fish in relation to its hunting behavior, although the prey must view the front end of the leaf fish (pers. observ.). The side view of *Monoacanthus* in Cott shows the pale interorbital region, although he failed to mention it. Upon hearing the presentation of the paper in hand, G. S. Myers ventured the suggestion that this leaf fish may be so thin that the split-head pattern operates perfectly well in *Monoacanthus*. In fact, when it feeds the head may blacken around the interorbital stripe, increasing the split-head effect, while the color of the rest of the body may remain relatively unchanged.

I have observed in a related nandid fish, *Nandus* sp., that a split-head pattern appears during feeding. *Polyacanthus abbreviatus* Boulenger, is an

African nandid fish and a species that looks much like *Polyacanthus*; it also manifests the split-head pattern when feeding (G. S. Myers, A.S.I.H.).

A further example is the serranid fish *Promicropus lawoofatus* (Bloch). The young are black with yellow blotches and have a pronounced split-head marking. This species has been characterized as a lurking predator (Cluapay 1963, fig. 2).

Dermatolepis punctata Gill is another serranid fish, having a split-head pattern (fig. 7). Hobson (pers. comm.) observed that this grouper is secretive by day but that it comes out to feed during the crepuscular periods. "It is usually active close to the bottom during these periods, yet it not infrequently ranges up into midwater. Although I have not observed this fish feeding, it is a sluggish swimmer with a manner that certainly suggests it to be a stalking predator." As in the case of *Promicropus*, the split-head pattern is more evident in the smaller fish (less than 350 mm).

Hobson has also written to me about another serranid fish with a striking split-head pattern, the soap fish *Rypocara bicolor* (Valenciennes). It too is a stalking predator that is active primarily during the crepuscular or nocturnal periods, eating mainly benthic crabs and crustaceans, although some fishes are taken.

Still another marine fish showing the split-head pattern is the giant kelpfish, *Heterostichus rostratus* Girard (Hobson 1965, figure on p. 510). Hobson (pers. comm.) found that "*H. rostratus* stalks small crustaceans and fishes, moving very slowly and not darting forward for the capture until within a few inches of its prey."

The sternarchid fish *Aparomene albigera* (Linnaeus) is a velvety black animal having a creamy white stripe running dorsally from the snout back about one-third of the body length (pers. observ.). It is nocturnal, perhaps crepuscular in habits, feeding on small fishes, shrimp, and insect larvae (Walker 1965).

These examples, coupled with the observations on *Polyacanthus* and *Nandus*, led me to believe that fishes with a disruptive interorbital stripe might be characterized collectively as lurking or slow-moving predators. This generalization, however, had to be modified in light of a report by Magnuson and Prescott (1966; my thanks to them for sending the unpublished manuscript).

Part of their study dealt with the bonito, *Sarda chilensis* (Cuvier), a schooling pelagic fish. When fed, the bonito manifests 9 to 10 vertical black bars on the body. More germane, a prominent yellow stripe appears extending from the tip of the snout to the base of the first dorsal fin. Nakamura and Magnuson (in Magnuson and Prescott) have observed nearly the same color change in the skip-jack tuna, *Euthynnus pelamis* (Linnaeus). In both of these oceanic species the bars and the interorbital stripe fade as the animals finish feeding.

The male bonito also expresses these body bars and the interorbital stripe when engaging with another male in lateral display, a hostile act, or when maneuvering to spawn with a female. It is important to note here that bonitos do not display frontally, probably because they cannot back up well.

Nakanishi, Magnuson, and Percott all endorse the idea that the primary signal value of this color change relates to hostile and spawning behavior. But they suggest it may have acquired the secondary significance of indicating the presence of food.

The bars may communicate something in a school of bonitos, whenever the message sent. That the interorbital stripe has signal value, however, is unlikely. Seen from the side, as it must be most of the time by other



Fig. 6.—*Pomacentrus littoralis*. Photo by H. Steene.



Fig. 7.—*Damnothia punctata* in its tank. Photographed by E. S. Steben near Los Angeles, California.

bonitos, it is a poor signal; its effect is only to flatten the head slightly. It must be viewed from in front to be of use, and only the prey regularly sees this aspect of the bonito. The split-head color pattern of the bonito probably serves to disrupt its frontal profile, just as it does in *Polyosteorhynchus*.

If this is so, then why does the interorbital stripe appear when the bonito is involved in social, nonfeeding behavior? The reason might be that this is the simplest solution for the bonito. Any major arousal could activate the color change, only one kind of change being necessary. Since the interorbital stripe is not seen well, it is of little consequence in intraspecific interactions.

Now in *Polyosteorhynchus* a similar situation prevails. When spawning they develop the interorbital stripe, strikingly so in the female. This, too, could be accounted for by general arousal. But when two males fight, the inter-

orbital stripe is lacking, the space being darkly colored as in the rest of the "face."

The explanation is that *Polycomus* engages in frontal displays when aggressive. A distinctive intraspecific signal, in this case the head plus extended opercles, should be relatively solid in color to emphasize the profile, and/or have an uninterrupted profile. An interorbital stripe would work against this because of the splitting of the head and disruption of the profile.

The spawning female, who must swim toward the male, takes advantage of the interorbital stripe to avoid the hostile signal that the uninterrupted face conveys. Similarly, the various female toothcarps of the genus *Cyprinodon* show a split-head color pattern in the context of sexual or aggressive behavior (R. K. Liu, pers. comm.).

Another indication of the role of arousal lies in the closer observation of feeding behavior. It is only when the prey is difficult to capture, or the *Polycomus* very hungry, that the interorbital stripe appears. When they have been fed small fish a few days in a row, the stripe is turned on as the feeder enters the aquarium room. But lest the point be made too strongly, recall that arousal involving aggressive behavior inhibits the appearance of the interorbital stripe.

Thus the contrast-rich interorbital stripe, the split-head color pattern, seems to characterize certain predatory fishes, particularly those lying in ambush. In some it occurs only when foraging. In others it is a fixed part of the color pattern.

This discussion of the split-head color pattern would be incomplete if I did not point out some exceptions. There are fishes possessing such a color pattern that one would not ordinarily conceive of as being predators. One of the best examples is presented by the browsing marine angel fishes. Henry Fiddler (A.S.I.H.) has pointed out to me this remarkable exception, based on his studies of these fishes in the Caribbean Sea. In *Pomacanthus paru* (Bloch) yellow encircles the mouth, then runs up from the circle through the interorbital space back onto the nape, reaching the insertion of the dorsal fin; the background color is dark brown or blue. In *P. arcuatus* (Linnaeus) there is also a yellow circle around the mouth; similarly a stripe runs up from the circle between the eyes and back to the insertion of the dorsal fin; but the stripe also extends ventrally along the chin. Juvenile fishes of this genus feed on sponges and algae; they are blue with a yellow split-head pattern. When either the juveniles or adults are approached they tend to face the intruder. Thus while they may not be using the split-head pattern as a concealment device to allow them to approach prey, they may be using it to permit them to look directly at a potential predator while making themselves more difficult to locate visually.

I have observed another interesting exception in the loach *Betta kawai*

Smith. This is a pale-colored fish with a black line running from the tip of the snout back along the entire dorsal surface, curving down along the insertion of the caudal fin, and terminating at the ventral surface of the caudal peduncle (Stein 1962, fig. 452). Seen from in front, the fish has a split-head pattern. Seen from the side, it has a black line at the base of the short dorsal fin and a black bar along the base of the caudal fin. The habits of the fish could be summarized as follows: It nibbles on plants and along the bottom, probably finding its food largely by virtue of the taste buds on the small barbels around the mouth. Prepared commercial fish food is taken visually, however, as it falls through the water. It is a fish that might be characterized as an active, nervous swimmer, but it is not a lurking predator. On the other hand, it frequently chases and apparently disturbs large fish in an aquarium. It is also vicious in the way it persistently attacks members of its own species. Conceivably, it could be ripping mucus from large fishes, although I doubt this. Perhaps more significant, the snout is long and relatively flat. The fish is mostly on or near the bottom. Thus the split-head effect could be part of a more general pattern which is to bisect the fish altogether when seen from above. (*Asabote dorei* Gill is a dark fish with a pale middorsal stripe running from the back of the head to the tail; living just under the water surface, it would often be seen from above by potential predators such as birds.) The black at the insertions of the dorsal and caudal fin of *B. kawai* probably serves in species recognition.

The pomacentrid fishes of the genus *Ampiprion* do not fit the scheme of a lurking predator. While carnivorous, they tend to hover, dart and turn, occurring together in small groups in the open. The clue is that the several species differ most immediately in the distribution of a few bold white marks on an orange to brownish background. The number of possible placements of these few white marks on the sides of the bodies of the species must reach some limit set by the abilities of the species to distinguish them. Having a white stripe in the middorsal area increases the number of possible combinations. Since members of the species move slowly, hovering and facing one another, recognition from the front presents no problems. A parallel pattern of species-specific markings can be seen in the white discs on the foreheads of pomacentrid fishes of the genus *Dascyllus*; they are remarkably similar in behavior.

Two further comments need to be made. The first is to point out that in three serranid fishes (*Grammistes aulivarius*, *Pomoxis maculatus*, and *Dorosaurus punctatus*) the young animals have the split-head pattern while it is lacking in the adults. Knowledge of any associated change in feeding habits would illuminate the significance, or lack of it, of the split-head pattern in these fishes.

The final comment is to draw attention, though fleetingly, to the problems of phyletic and ecologic correlations. Why is it that some kinds of ambush

predators are not known to show the split-head pattern? A good example is the large genus *Sphaeramia*. I suspect this is simply another example of different phylogenetic groups having different solutions to the same problems. Also, other considerations may prevail, such as species recognition. It is also my impression that the split-head pattern is more widely distributed among marine fishes than among freshwater fishes. This may be no more than a consequence of the great species diversity of the tropical shore fishes.

In conclusion, whenever the typical this splitting coloration is seen, one can probably be safe in assuming first that it is for the purpose of concealment. When this is on the front end of the fish, chances are that the fish is a predator, and more likely of the lurking, ambush type than an active feeder such as the boxfish. In some instances, a fish will have a split-head color pattern in order to observe a potential predator while remaining unseen. In a few species it may even be important as an intraspecific signal. And finally, some fishes, and particularly those that tend to feed on the bottom or near the surface in open areas, may have a dorsal line splitting them along their length to better conceal them.

REFERENCES

- ARL, E. F., 1964. *Evolutionäre und phylogenetische Zusammenhänge zwischen Mollusken, Nematoden, Platyhelminthen und Anneliden*. Zool. Jb. Anat. 51: 205-222.
- BALLOU, G. P., and J. M. BALLOU-FAN ROON, 1956. Introduction to the ethology of cichlid fishes. *Behavior*, Suppl. 1: 1-242.
- BALLOU, G. P., 1962. Underground concealment as a method of predation. *J. Mamm.* 42: 421-424.
- BALLOU, G. P., 1963. Ethology of the Asian teleost *Budi badii*. I. Locomotion, orientation, aggregation and flight. *Trans. III. Inst. Acad. Sci.* 34: 175-186.
- 1967. Social behavior of a South American leaf fish, *Polycottus schmidti*, with an account of recurring pseudo-female behavior. *Am. Midl. Nat.*, in press.
- CLARKE, P., 1965. *Prosimops lineatus* (Bleeker). *Aquar. u. Terrar.* 10: 207-204.
- COTT, H. B., 1940. Adaptive coloration in animals. Methuen, London, 509 pp.
- HANNON, E. S., 1965. Forest beneath the sea. *Animals (London)*, 7: 506-511.
- GRONMAN, E. J., 1966. A taxonomic study of *Eux americanus* and its sub-species in Eastern North America. *Copeia*, 1966: 1-20.
- LEWIS, W. H., and S. F. HILDEBRAND, 1948. Systematic catalogue of the fishes of Terrigena, Florida, with observations on color, habits, and local distribution. *Pub. Terrigena Lab. Carnegie Inst.*, 54: 1-131.
- MAURINON, J. J., and J. H. PINCOFF, 1966. Courtship, locomotion, feeding, and miscellaneous behavior of Pacific boxfish, *Acanthaluteres oblitus*. *Anim. Behav.* 14: 54-62.
- REICHARDT, J., 1929. *Fauna Boreali-Americana*. Murray, London, 500 pp.
- ROOSELY, T. C., 1955. *Wonders of the Great Barrier Reef*. Angus and Robertson, London, 195 ed., 281 pp.
- SCHRE, J. L. B., 1950. The sea fishes of Southern Africa. *General News Agency, Capetown*, 555 pp.
- STARR, G., 1962. *Freshwater fishes of the world*. Longmans, London, 678 pp.
- WALKER, R., 1965. Black ghost fish. *Aquar. Jour.* 36: 452-454.



Fig. 1—A long-nosed hawkfish, *Oxyurichthys typus*, found in Hawaii. This specimen was photographed in an aquarium after being captured alive at a depth of 120 feet. Many *Oxyurichthys* exhibit this characteristic pose of resting on the pectoral fins. Photo by E. A. Stearns, Jr.

A RARE HAWKFISH *OXYURICHTHES TYPUS* BLEEKER FOUND IN HAWAII

ROBERT A. MOORE and DONALD E. MOORE¹
New York Aquarium, Dept. of Land & Natural
Science Laboratories of Resources, Division of
Marine Sciences, Fish & Game,
Brooklyn, N.Y. Honolulu, Hawaii

Two specimens of the long-nosed hawkfish *Oxyurichthys typus* Bleeker, never collected previously in Hawaii, were obtained in July, 1961. These two air-breathing fish were collected alive from 120 feet of water off Maui Point, Oahu, Hawaii. They were observed resting on the branches of a "black coral" tree (*Acropora grandis* Verrill (Class Anthozoa, Subclass Scleractinia, Order Scleractinia)).

The genus *Oxyurichthys* previously had been divided into three species, *Oxyurichthys typus* Bleeker, *O. morrisi* Fowler, and *O. yfumi* Böhlke and Briggs on the basis of color pattern, preopercular serrations, pectoral ray combinations, and comparative lengths of spines. Randall (1965), however, resolved the genus to a single species, *O. typus*, the type locality of which is Amboina, East Indies. Other localities are Sumatra; Philippines; Mauritius; Baja, California; Panama; and Gorgona Islands, Colombia. The two speci-

¹ Our gratitude is extended to Dr. John E. Randall, Director of the Oceanic Institute for his suggestions concerning the preparation of this manuscript.

nium collected (and here reported) by the authors, therefore, represent not only the first record for the Hawaiian Islands but for all of Oceania.

Taxonomic notes of the two Hawaiian specimens are presented below:

Specimen I (Bernice P. Bishop Museum No. 5557)

Standard length 100 mm; sex female; dorsal rays X, 13; anal rays III, 7; pectoral rays right side ii, 7, v; left pectoral rays ii, 6 vi; second lower unbranched pectoral ray equal to snout; second lower unbranched pectoral ray longest on left side; first unbranched pectoral ray longer on right side; lateral-line scales 51 or 52; large scale rows above lateral line 4; large scale rows below lateral line 10; tenth dorsal spine much shorter than 3rd dorsal spine; second anal spine equal to postorbital part of head and $\frac{1}{2}$ eye; preopercular serrations 20 left side, 25 right side. Body length 4.2 in standard length; snout length 1.9 in head length.

Specimen II (Bernice P. Bishop Museum No. 5557)

Standard length 83 mm; sex undetermined; dorsal rays X, 13; anal rays III, 7; pectoral rays ii, 7, v (both sides); lateral-line scales 51 or 52; large scale rows above lateral line 4; large scale rows below lateral line 10; tenth dorsal spine much shorter than 3rd dorsal spine; second anal spine equal to postorbital part of head and $\frac{1}{2}$ eye; preopercular serrations 16 left side, 18 right side. Body depth 3.8 in standard length; snout length 1.8 in head length.

The taxonomic characteristics of the Hawaiian specimens fall within the range of *Oxyurhinus typus*. Variations were noted in the number of preopercular serrations between the right and left sides of both fish and also between the two specimens themselves. The combination of branched and unbranched pectoral rays differed also between the right and left side in specimen number I. This demonstrates that these characteristics cannot be relied on to distinguish one species from another.

Oxyurhinus typus is apparently a deep-water tropical species living in waters ranging from 35 to 118 feet in depth. Four other specimens have been observed in Hawaiian waters. Brock, Fujimura, and Takata observed a specimen resting in black coral in 90 feet of water off Lahilahi Point, Oahu in 1957. This is probably the earliest sighting recorded for the species in Hawaii. Three specimens have been sighted since then in 180 feet of water off Waikiki, Oahu. Two of these were observed resting on the branches of "black coral" trees and a third was living within the wreck of a small boat. The Hawaiian observations indicate that this long-nosed hawkfish has an affinity for the black coral growth either for protection or concealment.

LITERATURE CITED

RANDALL, JOHN E., 1963. Review of Hawkfishes (Family Cirrhitidae). Proceedings of the U.S. National Museum, 118:3472:389-412.

SULFAMERAZINE RESIDUES IN TROUT TISSUES

ROGER LEE HANSON and PAUL E. DOUTERM

Bureau of Sport Fisheries and Wildlife
Eastern Fish Disease Laboratory
Lectown (P.O. Kearneysville), West Virginia 25430
and
Nevins State Fish Hatchery
Madison, Wisconsin 53711

In the trout industry, sulfamerazine has been used primarily to control furunculosis. However, it is useful as a prophylactic treatment against kidney disease (*Corynebacterium*) and has been reported to be of help in the control of some *Aeromonas*, *Pseudomonas* and *Gyrodactylus* infections.

A review of the literature concerning the use of sulfamerazine for the treatment of disease in trout revealed a lack of the tissue residue data necessary for approval of the drug by the Federal Food and Drug Administration (F.D.A.). Smieszko and Fridde (1951, 1952) have shown that sulfamerazine concentrations drop rapidly in trout tissues when the drug is removed from the diet. However, these studies were not carried much beyond the point where the drug concentration fell below the minimum therapeutic level. Therefore the Eastern Fish Disease Laboratory and the Wisconsin State Conservation Commission conducted a series of experiments to fill this gap in the information required by the F.D.A.

METHODS

Rainbow, brook, and brown trout were randomly selected from the production stocks at the Lectown and Bowden National Fish Hatcheries (W. Va.) and the Nevins State Fish Hatchery (Wisconsin).

Rainbow and brook trout tests were carried out at 54 F. constant temperature (Lectown) and 45 F. average temperature (Bowden). Tests with brown trout were carried out at 50 F. constant temperature (Nevins) and 45 F. average temperature (Bowden) (table 1).

Sulfamerazine was fed in pelleted diets to all test fish except the Wisconsin brown trout. These fish had to be put on a medicated meal-meal mixture when they refused to eat the medicated pellets. The medicated pellets were prepared by coating commercial pellets with a slurry of sulfamerazine in hot (120-140 F.) 4% gelatin-water. The dosage rate was 10 grams per 100 pounds of fish per day. The medicated food was fed for 14 consecutive days with the exception of one lot of Wisconsin brown trout fed only 12 days and the brown trout at the Bowden National Fish Hatchery which were

TABLE 1. A listing of the hatcheries where tests were conducted, the species of fish and their age, and the water temperature.

	Tax number	Species	Water temperature	Age (months)
Eastern Fish Disease Laboratory, Leeches, West Va.	1-5	Rainbow trout	54°F. (12°C.) constant	15
		Brook trout		15
	1-8	Rainbow trout	54°F. (12°C.)	18
Bowden National Fish Hatchery, Bowden, West Va.		Brook trout		18
	1-8	Rainbow trout	45°F. (7.7°C.) average	17
		Brook trout		16
Nevis State Fish Hatchery, Madison, Wisconsin	1	Brown trout	50°F. (10°C.) constant	8
	2	Brown trout		8
	3	Brown trout		10
	4	Brown trout		19

fed on a three-days-on and three-days-off schedule during the 14 days. All feeding was on a free choice basis.

Controls fed non-medicated diets were carried in all tests.

The tissue residue analyses were done by one of the following methods depending on the expected amount of drug and the laboratory doing the analyses.

Modified Bratton and Marshall clinical method (1939)

The tissue is extracted with and diluted 1:20 with water, the protein precipitated with trichloroacetic acid, and color developed with N-(1-Naphthyl)-ethylene diamine. Because the sample is diluted only high tissue concentrations of drug can be measured by this procedure. Values below 10 ppm would be questionable.

Modified Goth clinical method (1942)

This is a modification of the Bratton and Marshall method. Acetone replaces the water for extraction of the drug. The use of acetone allows concentration of the sample and, consequently, analysis of lower tissue concentrations is possible.

Modified Mooney and Pasarela analytical method (1964)

This method employs an ion exchange column to remove interfering tissue constituents which react similar to sulfonamides. Color is developed as in the Bratton and Marshall method. A minimum of 2.5 micrograms of a

sulfonamide can be accurately determined. However, a 25-gram sample is necessary.

RESULTS

The several experiments, reported here, provided data from rainbow and brook trout at two temperatures but only one age group. The brown trout experiments provided two age groups in addition to the two temperatures.

As a rule, in all three species, the highest levels of sulfamerazine were found in the liver, with kidney, blood, and muscle following in that order. These results are similar to those reported by Sniezko and Friddle (1951, 1952) although they found the blood levels to be slightly higher than the kidney levels.

The three trout species presented a very similar pattern of drug elimination. Withdrawal of the drug from the diet resulted in a rapid decrease of the drug levels in all tissues to a relatively low range of values within 7 to 10 days. However, residues were present for a considerable length of time thereafter (figs. 1, 2, 3).

The comparison of figures 1, 2 and 3 indicates that the rate of elimination of sulfamerazine from brown trout muscle was slower at 45°F. than at 50°F., but indicates little difference in the rates of elimination from rainbow and brook trout muscle at 45°F. and 54°F.

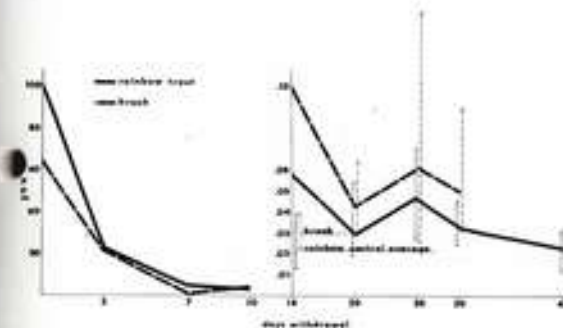


Fig. 1.—Average and (two \pm three) sulfamerazine levels in the muscle of rainbow and brook trout post treatment. Water temperature 54°F. (12°C.). The vertical bars indicate the range of values. Test 1-8 Eastern Fishery.

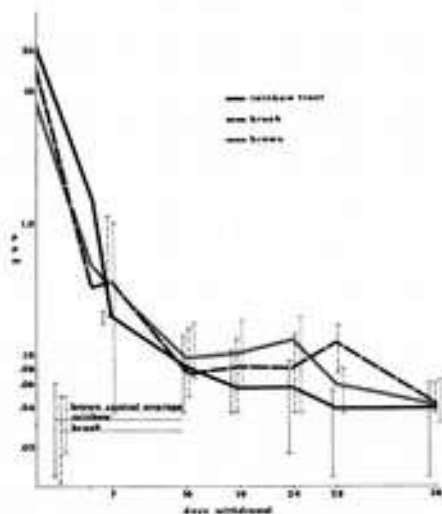


Fig. 2.—Average total (free + bound) sulfamerazine levels in the muscle of rainbow, brook and brown trout post treatment. Water temperature 43^o (11.2^oC) average. The ordinate scale is logarithmic. The vertical bars indicate the range of values. Test 1-8 Sweden Hatchery.

Rainbow trout and brown trout acetylate (bind) only small amounts of sulfamerazine whereas the brook trout acetylate large quantities, especially in the liver (table 2). The levels of acetylated (bound) drug drop as rapidly as the free drug levels after withdrawal of the drug.

DISCUSSION

Broadcast feeding (free choice) results in uncontrolled consumption of feed and, hence, drug. This, plus the probable uneven distribution of drug in the feed results in a wide range of drug levels among individual fish at any one time.

The validity of the temperature effect on the rate of elimination of sulfamerazine from brown trout muscle shown by the data is open to question.

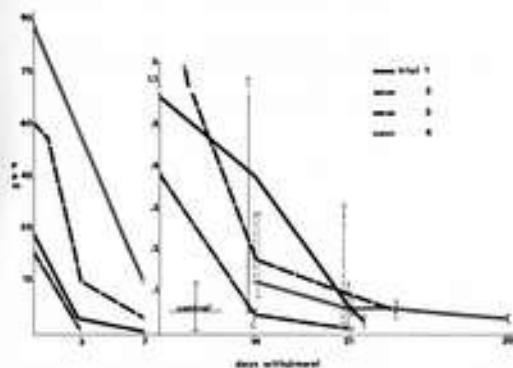


Fig. 3.—Average total (free + bound) sulfamerazine levels in the muscle of brown trout post treatment. The vertical bars indicate the range of values. Tests 1, 2, 3, 4 Maine Hatchery.

This difference in the rates of elimination may have been the effect of age rather than temperature. One generally assumes that, within limits, the younger a fish the higher is its rate of metabolism. Theoretically, the rate of metabolism governs the rate of drug elimination. The age of the brown trout used in these two experiments differed by seven months. This age difference may have been sufficient to cause a significant difference in the rate of metabolism and thus drug elimination.

The procedures used at high levels in the two tests for the tissue analyses were not identical. Recovery data (tables 3, 4) from samples of muscle with a known drug content indicates that the modification used in the Wisconsin tests had a lower recovery rate (average 72%) than the method used in the West Virginia tests (average 82%). This apparent difference in accuracy may have contributed to the difference in elimination rate shown.

Figures 1 and 2 show an increase in muscle residues for all three species at both temperatures about 25 days post treatment. This could represent a procedural error but tests 1-8 and 1-9 were analyzed six months apart. Identical errors are unlikely. This apparent increase probably represents

TABLE 2. Percentage of total sulfamerazine acetylated (bound).

Species	Blood	Tissue		
		Liver	Muscle	Kidney
Rainbow	11.06	35.55	28.40	14.93
Brook	32.55	60.26	23.19	33.59
Brown	2.52	11.66	2.96	6.77

TABLE 3. Validation of methodology, recovery of sulfamerazine from fortified rainbow trout muscle samples. (Modified Mooney-Pasarella performed by the Diablo Laboratories.)

ppm Added Sulfamerazine	ppm Total Sulfamerazine Recovered	Percent Recovery
0.05	.035	70
0.05	.047	94
0.10	.090	90
0.10	.072	72
0.20	.170	85
0.20	.176	88
1.00	.835	83
1.00	.799	80
		Ave. 82.75

TABLE 4. Validation of methodology, recovery of sulfamerazine from fortified brown trout muscle samples. (Modified Mooney-Pasarella performed at Nevins State Fish Hatchery.)

Micrograms Added Sulfamerazine	Micrograms Total Sulfamerazine Recovered	Percent Recovery
10.0	8.15	81.5
20.0	12.96	64.8
30.0	21.20	70.9
		Ave. 72.4

the release of drug from a small storage pool in the visceral fat. Unfortunately, this could not be verified since the visceral fat was not analyzed due to the sampling difficulties involved. Plasma protein bound drug is not a likely source since such binding is not particularly strong and the data (table 2) show only brook trout bind a significant amount of sulfamerazine. The liver does not appear to be a source since composite liver samples presented a similar increase.

The fact that the sulfamerazine levels in the blood and the kidney drop so rapidly as the levels in the other tissues leads to the speculation that sulfamerazine may be eliminated primarily across the gills of trout rather than through the kidneys as in mammals.

In brief, it was found that:

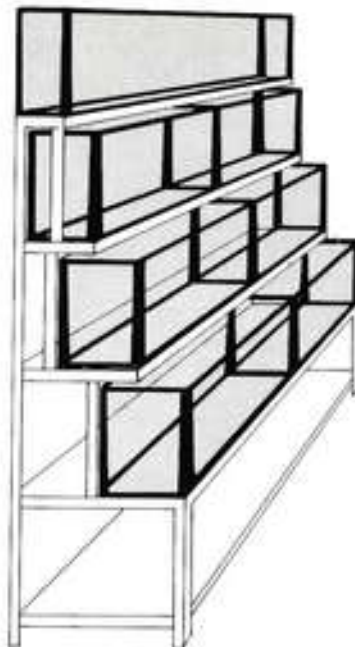
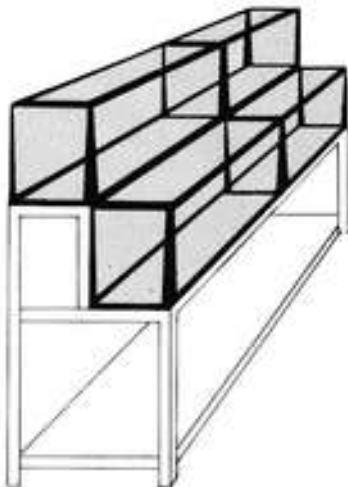
Sulfamerazine residues approach zero in rainbow trout muscle between 20 and 30 days after withdrawal of the drug from the diet.

Sulfamerazine residues approach zero in brook trout muscle between 30 and 40 days after withdrawal of the drug from the diet.

Sulfamerazine residues approach zero in brown trout muscle between 20 and 40 days after withdrawal of the drug, depending on the temperature. This is assuming the temperature difference shown by the data is valid. If it is not, zero residue may be reached between 20 and 30 days as with the rainbow trout.

LITERATURE CITED

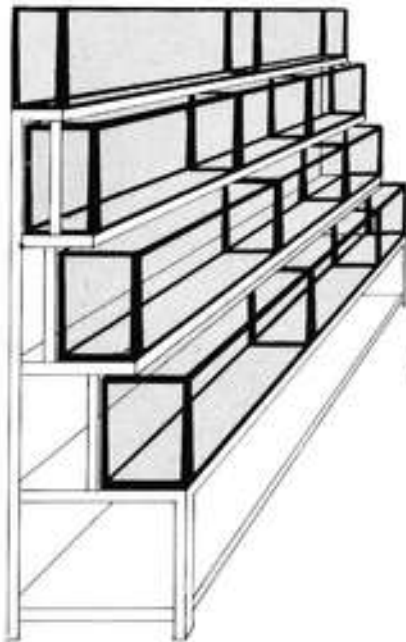
- BRATTON, A. C., and E. K. MARSHALL, 1939. A new coupling component for sulfamide determination. *Journal Biological Chemistry*, 128: 937-950.
- GOFF, A., 1942. A simple clinical method for determining sulfonamides in blood. *Journal of Laboratory and Clinical Medicine*, 27 (8): 827-829.
- MOONEY, R. P., and N. R. PASARELLA, 1964. A colorimetric procedure for the micro-determination of sulfonamides in animal tissues. *Agricultural and Food Chemistry*, 12: 125-127.
- SUTHERO, S. F., and S. B. FREEDLE, 1951. Tissue levels of various sulfonamides in trout. *Transactions American Fisheries Society*, 80: 240-250.
- 1952. Further studies on factors determining tissue levels of sulfamerazine in trout. *Transactions American Fisheries Society*, 81: 101-110.



**GIVE US A
ROUGH IDEA
OF YOUR LABORATORY
AQUARIUM
INSTALLATION NEEDS**

**WE'LL NOT ONLY
PROVIDE THE EQUIPMENT,
WE'LL MAKE SUGGESTIONS
THAT WILL
OFTEN IMPROVE
UPON YOUR IDEA
AND SAVE
YOU MONEY TOO.**

*Bader manufactures
a complete line
of quality aquariums
and aquarium
equipment*



BADER INDUSTRIES, INC., CHANGE BRIDGE ROAD, PINE BROOK, N.J.

THE DEATH VALLEY FISHES—
AN ENDANGERED FAUNA

MARTIN R. BRITTAN

One of the features of modern times has been the widespread alteration, replacement, extirpation, and extinction of existing native biotas through the direct and indirect activities of man. Countless examples may be found among fish faunas, resulting, as with terrestrial faunas and floras, from simple human predation (hunting, fishing, or trapping), destruction of the habitat (irrigation, farming, logging, construction work), or introduction of competing species, either domesticated (sheep and cattle) or wild (game and non-game birds, mammals, and fishes).

Some of the examples are well known, such as the destruction of the virgin Great Lakes forests by overlogging; others are obscure, such as the present case. Some are widespread, occurring over vast areas, such as the near extinction of the bison; others are limited to small areas, as with the Everglades Kite, or the present case. Some are complex, such as the decline of native fishes, both primary and anadromous, in the waters of the Sacramento-San Joaquin river systems; others are relatively simple, as in the present case: the threatened total extinction of five species of cyprinodont fishes in the Death Valley region of California and Nevada (and of a subspecies of a more widely spread cyprinid species).

The "Death Valley ichthyofauna", as we may call this assemblage, is limited to three species of *Cyprinodon* in the Amargosa River drainage of Ash Meadows and Death Valley, two species of *Empetrichthys*, one in Ash Meadows and one in adjacent but unconnected Pahrump Valley, and the cyprinid, *Rhinichthys osculus nevadensis* in Ash Meadows.

Empetrichthys is apparently a relict genus, perhaps derived in late Miocene or early Pliocene times from a species of *Fundulus*, while *Cyprinodon* probably penetrated into the area from the Colorado River drainage during the Pliocene. It is presumed most of the speciation has been Pleistocene and recent. *Rhinichthys* may have entered from the Colorado basin during Pluvial times. The taxonomic and zoogeographic aspects of the Death Valley ichthyofauna have been dealt with by Miller in a classic study (1948, 1950).

The species in question are normally threatened by the progressive dessication of bodies of water in arid climates; the lake and river systems in this part of the world have steadily shrunk since the last Pluvial period, and the forms are probably "on borrowed time" as it is, so it seems a double rime that man is hastening their extinction a hundred-fold.

Miller (1961: 394-8) gives seven factors in the depletion and extinction of the fish fauna of the southwest: (1) destruction of vegetation, (2) irrigation and dams, (3) mining operations, (4) depletion of ground water, (5) intro-



Fig. 1—Deep Spring on Tubbs Ranch, Ash Meadows, Nevada.



Fig. 2—Pond at Tubbs Ranch, Ash Meadows.

duction of exotics, (6) chemical treatment, and (7) changing climate. Industrial pollution is discussed under mining operations; it and pollution by domestic sewage, pesticides, and similar agents are bound to increase alarmingly in the future. The Death Valley ichthyofauna has been affected by only a few of these, but added to them must be the threat of vandalism.

In December, 1964, Dr. John Mizelle and I examined the status of the fish faunas in a number of springs, ditches, and creeks in Ash Meadows, Pahrump Valley, and Death Valley. In Ash Meadows we found that Deep Spring, on the property of Robert M. Tubbs, still had a good population of *Cyprinodon nevadensis mionectes*, as did the irrigation ditch leading from it, but in both environments the black molly, *Poecilia latipinna* (formerly known as *Molliensia latipinna*) was present. In the spring, on the day we examined it at least, the mollies outnumbered the pupfish about three to one; in the ditch there was about a 60-40 ratio in favor of the mollies, the *Poecilia* occupying the middle and upper levels, with the *Cyprinodon* occupying the bottom. Approximately two miles downstream where the irrigation ditch passes adjacent to Tubb's ranchhouse, the ditch possesses an additional exotic, *Gambusia affinis*, the mosquitofish, which far outnumbers both *Poecilia* and *Cyprinodon*; the mosquitofish had not, as of the date of our visit, migrated up the ditch to its source in Deep Spring. A farm pond immediately south of the ranch-house contained an extremely heavy population of *Gambusia*, a small population of mollies, and a very small number of *Cyprinodon*, in a rough proportion of 1000:20:1. There are now black bass in Crystal Spring (R. R. Miller, 1966, *in litt.*). A good population of *Cyprinodon nevadensis* survives below the main spring, and, according to James Deacon (1966, *in litt.*), the Nevada Dept. of Fish and Game has plans to eradicate the bass and allow the pupfish to reestablish themselves.

We did not examine other springs in Ash Meadows, but we fear the eventual contamination of all of them. Mr. Tubbs informed us that the exotic fishes appeared about a dozen years ago in the case of *Gambusia* and five or six years later in the case of *Poecilia*; he was unable to tell us how they got into the ditch or spring. In his opinion, the pupfish are steadily losing ground, though less rapidly than at first; this is probably because the two exotic poeciliids are not actively competing in the bottom environment. It seems unnecessary to point out that the introduction of any bottom-adapted fish in such a limited environment may quickly wipe out the surviving reduced *Cyprinodon* population.

Deacon adds further that the molly was also in Point-of-Rocks and Deep Spring, and associated ditches, in 1961, and had reached Jackrabbit (Eagle) Spring by 1965, probably by human transference, since the outlet is so swift.

A representative of a hatchery in Santa Ana, California, was to have visited Ash Meadows in early April, 1966, to stock aquarium fish and make



Fig. 3—Seining in ditch leading from Deep Spring on Tubbs Ranch.



Fig. 4—Overflow ponds adjacent to Saratoga Springs, Death Valley National Monument, California.

preliminary plans for buying land to be used for a wholesale hatchery operation. Dr. Deacon contacted Nevada Fish and Game, who at least temporarily stopped the stocking, but the company apparently still plans to set up the hatchery.

Another endemic cyprinodont, *Empetrichthys merriami*, known only from Ash Meadows, may now be virtually extinct. We observed a single, goldfish or pale yellowish fish in Deep Spring, about the size of a large *Cyprinodon* and more elongate, but it was too deep for positive identification; however, we are certain that it was neither a pupfish, molly, mosquitofish, *Rhinichthys scutellus nevadensis* (of which we observed a single example), or goldfish or other familiar species. However, Deacon has trapped extensively in Deep Spring without turning up any *Empetrichthys*, especially in 1963, 1964, and 1965. Traps were very successful on *E. latos* in Pahrump Spring. Crayfish are probably predators on *Empetrichthys*, since both are generally bottom dwellers. Deacon has also seen crayfish take *Cyprinodon*. *Empetrichthys merriami* was apparently excessively rare even before the white man came to Ash Meadows, the last capture being in 1957 (Miller, 1950:158), but the introduction of exotics will likely hasten total extinction, even if this has not already occurred.

The population of *Cyprinodon nevadensis nevadensis* in Saratoga Spring near the southern border of Death Valley National Monument appears normal, but the presence of a half-used box of laundry detergent near the wooden dock would indicate that the chance of this spring being contaminated is not at all remote. The area is surprisingly heavily-used by tourists (generally several camping parties per night, concentrated near the spring), but because of its remoteness from monument headquarters is only infrequently supervised by Park Service rangers. The removal of *Ceratophyllum* by campers (swimmers) may be ecologically important. The *Cyprinodon* population in the overflow lake is high, fortunately.

Devils Hole, a deep isolated spring on the edge of Ash Meadows, is encompassed in a separated segment of the monument, and was at the time of our visit protected by a stout wire-mesh fence with a steel gate with a padlock and chain. The total population of *Cyprinodon diabolis* is restricted to Devils Hole, and fluctuates between perhaps 100 and 500 individuals. While the population was in good condition, numbering, we estimated, about 350 (in January, 1966, Miller estimated the population at about 200), there were evidences of human disturbance. The chain on the gate had been cut and several dozen photographic flashbulbs and tin cans floated in the water. A piece of sounding line arched into the depths. Several months later, two scuba divers drowned in Devils Hole. It is my understanding that since then the monument authorities have planned to roof over the entire area with heavy wire screening.



Fig. 5—Devils Hole (beyond sign), Death Valley National Monument, Nevada.



Fig. 6—Explanatory sign at Devils Hole.



Fig. 7—Devils Hole, Death Valley National Monument, Nevada.

The Devils Hole population could easily be destroyed by introduction of exotic species or by accidental or deliberate vandalistic contamination by pollutants. The Park Service is making great efforts to save this species, but any form existing in such small numbers in such a restricted environment is threatened sufficiently by nature without the presence of so potentially destructive a factor as man.

The population of *Cyprinodon salinus* in Salt Creek is probably safe from man, since there are no campgrounds nearby and the creek has an appreciable flow; however, a trail runs alongside, so there is always the possibility of accidental or deliberate contamination. The Pupfish Slough population is presently isolated and safe from man. However, both populations are threatened by long-term progressive aridity, since Salt Creek is alive for only a few hundred yards and is, in most places, only two or three feet wide and a foot or so deep, while Pupfish Slough, though several acres in extent, is only a few inches deep in most places.

In Pahrump Valley, where most of the cotton in Nevada is grown, *Cyprinodon* is absent. *Empetrichthys latos* is restricted to three localities in the valley, each supporting a distinct subspecies. *Empetrichthys latos latos*



Fig. 8—Entrance to Devil Hole.



Fig. 9—Salt Creek, Death Valley National Monument, looking downstream (north).

occurs in the main spring pool at the Manse (Bowman) Ranch, where the population was estimated at 200 by Deacon in April, 1965 (*in litt.*, 1966) and by Miller (*in litt.*, 1966) as "pretty good." This pool is heavily contaminated with goldfish, *Carassius auratus*, which had been introduced to the pool several years previously by ranch employees (according to the employees). The Manse Ranch situation is discussed in considerable detail by Deacon, Hubbs, and Zahvanec (1964: 304-6). Two other subspecies of *Desmocerichthys* are now extinct. *Desmocerichthys later palustris* had disappeared with the drying up of a spring-fed ditch on the Pahremp Ranch, and *Desmocerichthys later amarus* with the going-dry of Raycraft Springs. Both events were supposedly connected with well-drilling operations (verbal communication with Mr. Frank Warner, foreman of Pahremp Ranch, who informed me that both of the springs and the irrigation ditch went dry after another land-owner apparently accidentally drilled through an impervious bed underlying an aquifer; the owner at Raycraft Springs, however, told Deacon that he filled in the spring because, about 1957, he got tired of mosquitoes).

It appears to the writer that while accidental damage to fish faunas in the course of human activities of a "constructive" nature (such as the above

well drilling, irrigation, dam building, insect control, etc.) are unfortunate enough, but the accidental or deliberate introduction of exotics is especially deplorable. Such introduction represents gross thoughtlessness, to say the least, ranging on through irresponsibility to downright selfishness and malice. When done by a government agency, with intelligent forethought, should the results not be as planned, it can be considered as simply unfortunate; when done by individuals with no thought to the future, it can be regarded as completely irresponsible, and in some cases downright criminal.

The introductions of exotics fall mainly into the following categories: (1) introduction of game or forage fish, or fishes for insect control, by government agencies; (2) introduction of aquarium or pondfishes by aquarists or pond-keepers, either to avoid the necessity of destroying them or to create a natural stock for subsequent exploitation; (3) accidental escape into new waters from the place of initial introduction. The initial introduction of the mosquitofish may be due to either of the first two factors, that of the mosly is almost certainly due to the second (as is that of the goldfish).

Aquarists and live fish dealers are presently agitated over the action of fish and game authorities in several states in seeking and obtaining legislation making illegal not only the planting of exotic fishes except by authorized authorities (the fish and game agencies themselves), but also the importation and even the keeping of certain species. They are in large part responsible for their own dilemma, since in carelessly allowing exotics to

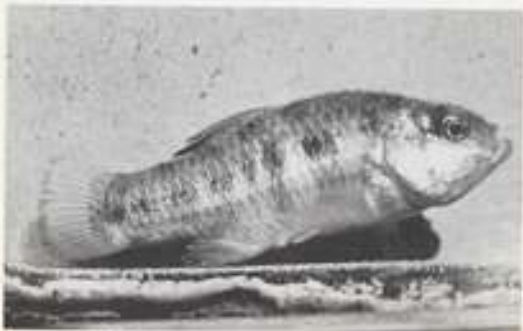


Fig. 10.—*Cymatodus n. nardoni*, male, from Saratoga Springs.



Fig. 11.—*Cymatodus n. nardoni*, female, from Saratoga Springs.



Fig. 12.—*Cymatodus diabolus*, male, the Devil Hole pupfish (note the absence of ventral fin in this species).



Fig. 13.—*Cymatodus* yellow, the Salt Creek pupfish, probably a subspecies female.

re in dumping them into native waters to avoid the trouble of properly rearing them, they are damaging native and long-existing introduced and forcing conservation agencies to react against the threat. There is full of examples of the introduction of exotics, and the results can be as often harmful as beneficial by human standards, but to the faunas affected the results have almost always been deleterious. In those cases where exotics are introduced into a large resilient fauna the results are serious, but where the introductions are into a depauperate fauna of highly specialized species with low ecological valences, the results can only be disastrous. The near-total depletion, extirpation, or loss of the existing fauna can be regarded as particularly tragic when the biogeography is as restricted and unique as is the fish fauna of the Death Valley region.

BIBLIOGRAPHY

- CLARK, JAMES E., CLARK HERRIN, and BERNARD J. ZIMMERMAN, 1964. Some effects of introduced fishes on the native fish fauna of southern Nevada. *Gophers* (2): 8, table 1.
- CLARK and JAMES E. DEACON, 1964. Additional introductions of tropical fishes into southern Nevada. *The Southwestern Naturalist* 9 (4): 249-51.
- DEACON, J. E., 1962. Fishes and fisheries of Nevada. *Nevada State Fish and Game Commission*, 782 pp.
- DEACON, J. E., 1946. Correlation between fish distribution and Pleistocene geology in eastern California and southwestern Nevada, with a map of the recent waters. *Trans. Geol. Surv.* 54: 43-53, figs. 1-2.
- DEACON, J. E., 1946. The Cyprinodont fishes of the Death Valley system of eastern Calif. and western Nevada. *Ann. Entomol. Soc. Amer.* 39 (1): 1-155, figs. 1-5, I-XV, maps 1-3.
- DEACON, J. E., 1946. Speciation in fishes of the genera *Cyprinodon* and *Epiplatys*, inhabiting Death Valley region. *Evolution* 4 (2): 155-63, fig. 1, table 1, maps 1-2.
- DEACON, J. E., 1946. Man and the changing fish fauna of the American Southwest. *Pap. Mich. Acad. Sci., Arts, and Letters* 46 (1960): 365-404, fig. 1.
- DEACON, J. E., 1946. Is our native underwater life worth saving? *National Parks Mag.* 37 (1): 1-9, 4 figs.
- DEACON, J. E., 1946. Extinct, rare, and endangered American fishes. *XVII International Year of Zoology (Washington) Science and Man Symposium* 8: 4-11, figs. 1-11.
- DEACON, J. E., 1946. The introduced fishes of Nevada, with a history of their selection. *Trans. Amer. Fish Soc.* 75: 173-93.



Fig. 1.—A black crappie, *Pomoxis nigromaculatus*. Photo by Al Giddings.

OBSERVATIONS ON THE FOOD HABITS OF ADULT BLACK CRAPPIE IN A CALIFORNIA LAKE

ROBERT N. LIA
Wildlife-Fisheries
Museum of Vertebrate Zoology
University of California, Berkeley

The stomach contents of 31 adult black crappie, *Pomoxis nigromaculatus*, were analyzed by the gravimetric and the frequency of occurrence methods outlined by Lagler (1956). These fish were collected by standard angling methods at Kelly Lake, 2 miles east of Watsonville, Santa Cruz County, during late March and early April of 1964. The fish varied from 155 to 260 mm in standard length and 141.9 to 496.9 g in weight, with mean values of 191.5 mm for length and 280.0 g for weight. All fish were in prime spawning condition. Coefficients of condition, *K*, were calculated for all specimens. An average *K* value of 3.36 indicated their excellent state, with values ranging from 2.77 to 3.88.

Of the 31 stomachs examined, 24 contained food items. Entomostraca were found in 21 of the 24 stomachs and composed 95.0 percent by weight of all food taken. Of the two organisms recognized as entomostraca, a cladoceran (*Daphnia* sp.) represented approximately 90 percent of the diet, and a copepod of the genus *Diaptomus* comprised the other 5 percent.

Only one form of insect was found in the stomachs, a chironomid (*Pelopia* sp.). Chironomid pupae were the second most important food item utilized, composing 4.3 percent of the total stomach content by weight, and found in 17 stomachs. Chironomid larvae formed only an insignificant part of the food taken, composing 0.1 percent of the total mass by weight.

The amphipod *Hyalina astuta* was found in three of the 24 stomachs and composed 0.2 percent of the total weight.

Plant material was found in four stomachs. However, it seems likely that this matter was ingested inadvertently.

A summary of the food contents of the 24 stomachs is given in Table I.

TABLE I

Stomach contents of 24 adult black crappie taken in Kelly Lake

Food Item	Frequency of occurrence		Percentage composition of food by weight
	f	percent	
Crustacea			
Entomostraca	21	87.5	95.0
Amphipoda	3	12.5	0.2
Insecta			
Chironomidae			
Pupae	17	70.8	4.3
Larvae	2	8.3	0.1
Plant matter	4	12.5	0.4

No definite conclusions can be drawn on the basis of so restricted a sample, but it may be significant that other food-habit studies of black crappie, made in their native range, indicate a similar preference for entomostraca and immature insects (Ewers, 1933; Neal, 1962).

The author is grateful to Robert P. Dempster, Steinhart Aquarium, California Academy of Sciences, for allowing the use of facilities for photographic work.

LITERATURE CITED

- EWERS, LILA A., 1933. Summary report of crustacea used as food by the fishes of the western end of Lake Erie. Amer. Fish. Soc. Trans., 61: 179-190.
- LAZAR, KARL F., 1936. Freshwater fishery biology. Wm. C. Brown Co., Dubuque, Iowa, 421 pp.
- NEAL, RICHARD A., 1962. White and black crappies in Clear Lake, Summer, 1960. Proc. Intern. Acad. Sci., 1961, 69: 257-253.

SUBSCRIBE
TO
ICHTHYOLOGICA
The Aquarium Journal



Please print. Detach and send to:

T.F.H. PUBLICATIONS, INC.
243 Conestoga Avenue, Jersey City, N. J. 07302

Gentlemen:

I am interested in subscribing to *Ichthyologica*, the Aquarium Journal. Enclosed please find check, money order or cash for:

Sample copy — \$1.00

12 issues (3 years) — \$10.00

12 issues at special student price — \$5.00

School Attending _____

Name _____

Street _____

City _____

State _____

Zip _____

NEW
THE THINKING MAN'S FILTER... THE MIRACLE HIPOWER FILTER*



- 1 The new Miracle HiPower Undergravel Filter is about 1" lower in the back than the older model and has optional brackets for screws to fit on either side.
- 2 Just apply pressure firmly with your thumb and the disc easily bends up to accommodate a Miracle HiPower stem assembly.
- 3 Don't break both discs unless you want to use both stem assemblies in one filter, as this kit contains only two filter stems, one for each filter.



- 4 Snap the filter stem into place; the wider nipple fits into the wider slot. In all cases the narrow air tube goes to the rear as shown.



- 5 This is the new Miracle HiPower Undergravel Filter set up and ready to be placed in your aquarium, where it never need be serviced again.



- 6 For ultra-high power filtration where your fishes are overcrowded, you can use two filter stems in one filter. (Perhaps stick and sell extra stems.)
- 7 The normal approach is a filter stem in each filter at opposite corners, so the stems "hide" in the corners of the aquarium.
- 8 Look for the Miracle HiPower Undergravel Filter in the red and black box at your petshop. It is the most powerful undergravel filter in the world!

*Patent Pending in the USA, Canada, England, Germany, Hong Kong and Japan.

MIRACLE PLASTICS (N.J.) CORP. BOX 33, JERSEY CITY, N. J. 07303
 MIRACLE PLASTICS CORP. BOX 9128, LONG BEACH, CALIF. 90810

AQUATIC FISHERIES HAS THE MOST MODERN WHOLESALE FISH FACILITIES IN THE WORLD / AQUATIC FISHERIES GUARANTEES YOU A FULL LINE OF IMPORTED AND DOMESTICALLY RAISED TROPICALS AT THE LOWEST POSSIBLE PRICES FOR QUALITY FISH / BEAUTIFUL LIVEBEARERS OUR SPECIALTY! IMPORTS ARE OUR BREAD & BUTTER!



Aquatic Fisheries

P.O. Box 16, Vero Beach, Florida 32960, Phone (305) 561-5277

Import-Export - WHOLESALE ONLY - Exotic Tropical Fish - Cable Address "Aquatfish"

Your name should be on the cover of this magazine



If it isn't, mail the coupon.

Subscribers receive every issue of TROPICAL FISH HOBBYIST without fail. Subscribers are kept informed about every important development in the aquarium hobby... the new fish, the new people, the new products, the new techniques... every month. They also know that they'll always be up to date with their free supplements to EXOTIC TROPICAL FISHES. Subscribe NOW to TROPICAL FISH HOBBYIST, the biggest (more pages, more pictures, more information, more readers), most colorful (a minimum of 24 color pages in each issue), most interesting aquarium magazine in the world.

TROPICAL FISH HOBBYIST
245 Cornelian Avenue, Jersey City, N. J. 07302

Enclosed please find check, money order or cash for:

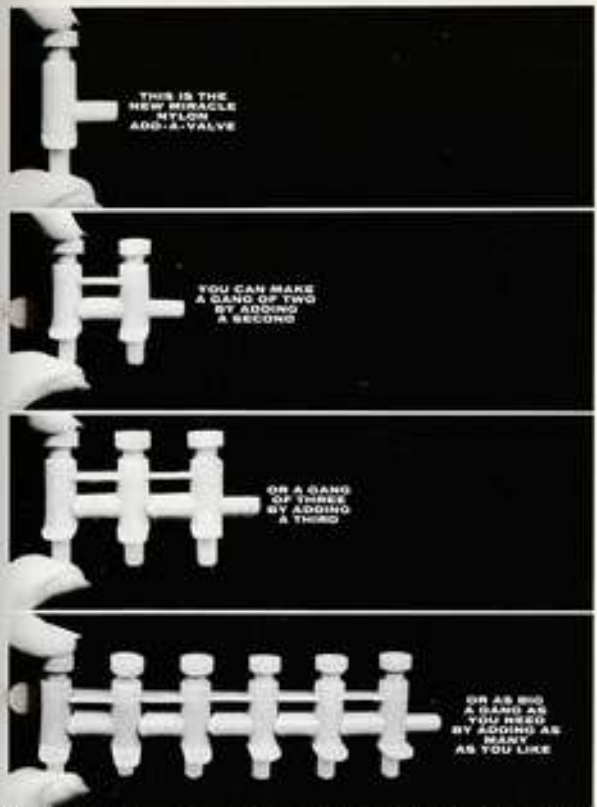
- Sample copy - 35¢
- 12 issues of Tropical Fish Hobbyist, \$3.50
- 24 issues of Tropical Fish Hobbyist, \$6.25
- 60 issues of Tropical Fish Hobbyist, \$13.00

Foreign Subscribers: Add 60¢ per year's subscription.

Name _____

Street _____

City _____



THIS IS THE NEW MIRACLE NYLON ADD-A-VALVE

YOU CAN MAKE A GANG OF TWO BY ADDING A SECOND

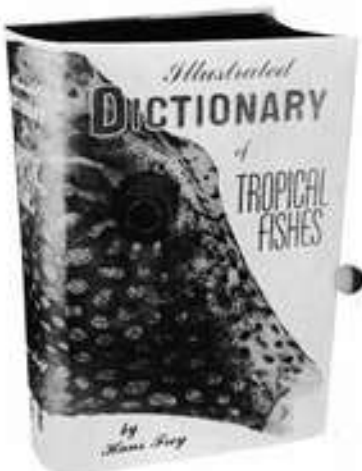
OR A GANG OF THREE BY ADDING A THIRD

OR AS BIG A GANG AS YOU NEED BY ADDING AS MANY AS YOU LIKE

With the revolutionary new Miracle Nylon Add-A-Valve, you get unlimited flexibility in creating custom-made gangs with exactly the number of air outlets you want. Only the specially made Miracle Nylon Add-A-Valve offers all these advantages: The Ability To Custom-Create Gang Valves - Absolute Airflow Control (No need for constant adjusting.) - Can't Introduce Metal Poisons To Your Aquarium - Less Expensive (Nylon, which does the job better than metal anyway, costs less.) - Guaranteed Corrosion-Proof. Buy the Miracle Nylon Add-A-Valve at your local pet shop.

MIRACLE PLASTICS (N.J.) CORP. BOX 33, JERSEY CITY, N. J. 07303
MIRACLE PLASTICS CORP. BOX 9120, LONG BEACH, CALIF. 90810

Hans Frey's
**ILLUSTRATED
DICTIONARY OF
TROPICAL FISH**
\$7.95



Never before in the English language has a dictionary of tropical fish appeared. This huge volume, translated from the German by Dr. A.

Viggo W. Schultz, brother of Harald Schultz, covers every fish, plant and snail that has ever reached any popularity in the last 50 years! It is illustrated with more than 1,000 photographs and drawings in color and black and white. There is no book like it that compares in size, scope, or completeness. In German it sold 18,000 copies in less than two years, making it a national best-seller. This book is a *must* for every beginner and advanced hobbyist alike who wants to identify a rare fish, plant or snail, or who needs information on any aquatic subject. Besides the subjects already mentioned, there are entries on breeding, water, infusoria, diseases, heaters, filters, pumps, feeding, live foods, fish collecting, marine fish, goldfish, cold water fish and terrarium life. Available now at your petshop. Add 50c. for postage and handling if ordered from the publisher. Send cash, check or money order to

Published by T.F.H. PUBLICATIONS, INC.
245 Cornelison Avenue Jersey City, N. J. 07302

100

**THIS IS A
BADGER
AQUARIUM-
TO BREAK IT
YOU HAVE
REALLY
GOTT TO TRY!**



BADER INDUSTRIES, INC., CHANGE BRIDGE ROAD, PINE BROOK, N.J.

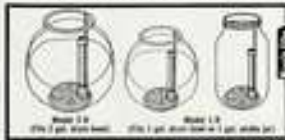


"The Little Miracle!"

FISH BOWL FILTER

Tropical Fish Breeders!... "The Little Miracle" 1-R filter is the perfect answer to caring better. This wonderful little filter now makes any ordinary glass fish bowl a "Sparkling Beauty Without Work" - Miracle Filter aquarium. No more cloudy, messy fish bowls to clean. **2-R** How keep live food alive... **3-R** *Capnia*, *Brine Shrimp*, *Red Worms*, **Marine Tropical Hobbyists!**... "The Little Miracle" fish bowl filter solves the annoying and costly problem of your *Saltwater* can be saved? **2-R** *C-G* See your favorite store now for the "The Little Miracle" filter. **2-R** Available in one and two gallon sizes.

2 NEW SIZES - MANY NEW USES



MIRACLE PLASTIC COMPANY, 201 1/2 Long Beach, Calif.

